



## FIT Transferrin NS-Prime Test

Catalogue No.	Product name
911319	FIT Transferrin NS-Prime
911320	FIT Transferrin NS-Prime Calibrator

Manufactured by:

**Alfresa Pharma Corporation**  
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European Authorized Representative

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**Refer to the following URL for the package inserts in languages other than English:**

<http://www.alfresa-pharma.co.jp/english/works/packageinserts.html>



### FOR *IN VITRO* DIAGNOSTIC USE ONLY

#### 1.0 INTENDED USE

**1.1 FIT Transferrin NS-Prime** is a reagent to quantitatively determine human transferrin concentrations in feces using the **Discrete Clinical Chemistry Analyzer NS-Prime**, which employs a colloidal gold immune colorimetric and turbidimetric method. This reagent is intended for colorectal cancer screening. FIT stands for fecal immunochemical test.

**1.2 FIT Transferrin NS-Prime Calibrator** is a calibrator to construct a calibration curve. Transferrin value in specimen is determined using the calibration curve.

#### 2.0 PRINCIPLES OF THE EXAMINATION METHOD

##### 2.1 Summary and explanation of the test

Fecal immunochemical test (FIT) is used to diagnose hemorrhagic gastrointestinal diseases. FIT is particularly useful for colorectal cancer screening. **FIT Transferrin NS-Prime** is a kit to measure human transferrin concentrations in feces using an immunochemical method combined with a colloidal gold colorimetric and turbidimetric method. This colloidal gold immune colorimetric and turbidimetric method is intended to measure an optical color change due to agglutination between colloidal gold-conjugated rabbit anti-human transferrin polyclonal antibodies and fecal human transferrin. This test is highly specific and sensitive.

##### 2.2 Principles of the test

The reaction of colloidal gold-conjugated anti-human transferrin polyclonal antibodies with human transferrin in feces produces a color change due to agglutination of colloidal gold particles through the antigen-antibody reaction. Human transferrin concentration in feces is determined by measuring the color change over time.

#### 3.0 TRACEABILITY OF VALUES ASSIGNED TO CALIBRATORS AND TRUENESS-CONTROL MATERIALS

Human transferrin is a reference material to calibrate the calibrator and controls. Human transferrin concentrations are determined using the IRMM ERM-DA470k / IFCC.

#### 4.0 COMPONENTS

##### 4.1 FIT Transferrin NS-Prime

Reagent 1 and Reagent 2 are combined and packaged into one set. A total of 300 tests can be conducted.

##### 4.1.1 Reagent 1 REAG11

Reagent 1 contains 33 mL per bottle. Each bottle contains:

MES Buffer	150 mmol/L
Sodium azide	< 0.1%

Reagent 1 contains less than 0.1% sodium azide. Attention, see **8.0 WARNINGS AND PRECAUTIONS**. Material Safety Data Sheet is available upon request by a professional user.

##### 4.1.2 Reagent 2 REAG21

Reagent 2 contains 13 mL per bottle. Each bottle contains:

TES Buffer	4.4 mmol/L
Bovine serum albumin	0.03%
Sodium azide	< 0.1%
Colloidal gold-conjugated antibodies	anti-human transferrin polyclonal antibodies 333 µL/mL

Reagent 2 contains less than 0.1% sodium azide. Attention, see **8.0 WARNINGS AND PRECAUTIONS**. Material Safety Data Sheet is available upon request by a professional user.

##### 4.2 FIT Transferrin NS-Prime Calibrator

Four Calibrator (lyophilized) vials and one Calibrator Solution bottle are packaged, permitting four times calibration.

##### 4.2.1 Calibrator (lyophilized)

Each vial is reconstituted with 1.0 mL of Calibrator Solution **immediately before use**. Each vial contains:

MES Buffer	3.2 mg/vial
Sucrose	10 mg/vial
Bovine serum albumin	1.0 mg/vial
Human transferrin	400-600 ng/vial

##### 4.2.2 Calibrator Solution

Calibrator Solution contains 12 mL per bottle. It contains:

MES Buffer	30 mmol/L
Sodium chloride	1.1%
Bovine serum albumin	0.15%
Sodium azide	< 0.1%

Calibrator Solution contains less than 0.1% sodium azide. Attention, see **8.0 WARNINGS AND PRECAUTIONS**. Material Safety Data Sheet is available upon request by a professional user.

#### 5.0 ADDITIONAL REQUIRED EQUIPMENT

##### 5.1 Analyzer

**Discrete Clinical Chemistry Analyzer NS-Prime**  
**Discrete Clinical Chemistry Analyzer AA01**

**FIT Transferrin NS-Prime** is only used with **Discrete Clinical Chemistry Analyzer NS-Prime**, but **FIT Transferrin NS-Prime Calibrator** is used with both **Discrete Clinical Chemistry Analyzers NS-Prime** and **AA01**.

##### 5.2 Specimen Collection Container

**Specimen Collection Container A**

##### 5.3 Control

**FIT NS-Prime Control**

##### 5.4 Specimen Diluent

**FIT NS-Prime Specimen Diluent**

**FIT AA01 Specimen Diluent**

**FIT Transferrin NS-Prime** is only used with **FIT NS-Prime Specimen Diluent**, but **FIT Transferrin NS-Prime Calibrator** is used with both **FIT NS-Prime Specimen Diluent** and **FIT AA01 Specimen Diluent**.

##### 5.5 Wash Solution

**NS-Prime Wash Solution**

**Wash Solution A**

#### 6.0 REAGENT PREPARATION

##### 6.1 FIT Transferrin NS-Prime

Mix the Reagent 1- Reagent 2 combined bottle thoroughly before each use, because colloidal gold particles may precipitate during storage. Mix it by slowly inverting to avoid creating bubbles. If bubbles occur, the instrument sensor will read an incorrect volume and the bottle will be deleted from the system. Pipette out any bubbles down to the level of the liquid before placing the bottle on board.

##### 6.2 FIT Transferrin NS-Prime Calibrator

Allow Calibrator and Calibrator Solution to reach room temperature. Reconstitute the lyophilized Calibrator with 1.0 mL of Calibrator Solution. Mix well by inversion before use.

#### 7.0 STORAGE AND SHELF LIFE AFTER FIRST OPENING

##### 7.1 Storage

All reagents are stored at 2–8°C. Do not freeze.

##### 7.2 Storage and shelf life after first opening

##### 7.2.1 FIT Transferrin NS-Prime

Once opened, use the reagent within one month and store at 2–8°C.

##### 7.2.2 FIT Transferrin NS-Prime Calibrator

Once opened and reconstituted, with the Calibrator Solution, use immediately. Store the Calibrator Solution between 2–8°C.

#### 8.0 WARNINGS AND PRECAUTIONS

##### 8.1 General precautions

For *In vitro* diagnostic use

Only experienced laboratory personnel should use this; the test should be used in a manner consistent with Good Laboratory Practice.

##### 8.2 Safety precautions

- Do not pipet by mouth.
- Some reagents contain less than 0.1% sodium azide. Upon exposure to the eye or skin or accidental ingestion, take emergency measures such as washing with plenty of water. Consult a doctor if necessary.
- Do not smoke, eat, or apply cosmetics in areas where patients' specimens or kit reagents are handled.
- Cuts, abrasions, and other skin lesions should be properly protected with an appropriate waterproof dressing.
- Take care to avoid self-inoculation, splashing to the mucous membrane, or generation of aerosols.
- Wear laboratory gloves while handling patients' specimens or disposing of solid or liquid wastes.
- Cautions upon disposal
  - Reagent 2 of **FIT Transferrin NS-Prime** contains 0.20 g/L ethylenediaminetetraacetic acid copper (II) disodium (27 mg/L as copper). Upon disposal, comply with relevant legal provisions.

- Some reagents contain less than 0.1% sodium azide. Sodium azide can react with copper and lead plumbing to form explosive metal azides. Regulations currently in use regarding dangerous waste elimination must be followed. If disposed in the sink, rinse with plenty of water.
- Upon disposal of reagents or other materials, comply with relevant legal provisions.
- Some reagents contain bovine serum albumin free from known infectious agents. However they should be considered potentially infectious and handled with care to avoid infection.
- All human specimens should be considered potentially infectious. Handle all specimens as if capable of transmitting HBV, HCV, HIV, or other microbes. Decontaminate and dispose of specimens and all potentially contaminated materials as if they contain infectious agents.
- Human origin raw materials contained in **FIT Transferrin NS-Prime Calibrator** are negative for HBs antigen, HCV antibody, and HIV antibody. However, handle them with care as they are potentially infectious. No known test method can offer complete assurance that products derived from human sources will not transmit infectious agents.

##### 8.3 Limitations

- Do not use reagent containers for purposes other than this test.
- Do not separate reagent containers that connect **FIT Transferrin NS-Prime**. Do not use them in combination with other reagent containers even if they are the same lot.
- Do not damage or stain the bar codes on the labels of each container.
- Do not replenish or mix reagents. Also, do not mix reagents of different bottles even if they have the same lot number.
- Do not use combinations of different lot numbers within the products.
- Use of reagents, disposables, or spare parts other than those supplied by the authorized distributor may produce incorrect results.
- Reconstitute the calibrator with the calibrator solution supplied. Do not use other solution.
- Do not recycle the bottle. It may be infectious.

#### 9.0 PRIMARY SAMPLE COLLECTION, HANDLING, AND STORAGE

Use human fecal specimens for measurement.

- Scrape the surface of fecal specimen with the stick of **Specimen Collection Container A** thoroughly to prepare a sample.
- Specimens whose concentrations exceed the upper limit of the calibration curve should be diluted with **FIT NS-Prime Specimen Diluent** and retested.

##### 9.1 Specimen collection

Sample feces onto the gutters of the collector stick by scraping several surfaces of fecal specimen. Place the stick into the collector body once only and fasten the stick tightly. See the instruction manual for **Specimen Collection Container A**.

##### 9.2 Specimen storage

The collected sample should be refrigerated between (2–8°C) until the sample can be tested. Once the sample is received for testing store between 2–8°C.

##### 9.3 Transferrin stability after feces sampling

The stability of human transferrin in the buffer of **Specimen Collection Container A** was examined. After storage of four different concentrations of human transferrin for 6, 13, and 35 days at –40, 7, 25, and 37°C, the residual ratios of human transferrin were as follows:

	–40°C, for 35 days	7°C, for 35 days	25°C, for 13 days	37°C, for 6 days
39 ng/mL	≥ 90%	≥ 90%	≥ 80%	≥ 70%
66 ng/mL	≥ 90%	≥ 90%	≥ 80%	≥ 70%
157 ng/mL	≥ 90%	≥ 90%	≥ 80%	≥ 70%*
296 ng/mL	≥ 90%	≥ 90%	≥ 80%	≥ 70%*

\* If residual ratios decreased to 70% or above, but the results were still positive (cut-off: 50 ng/mL).

This result is provided for reference only. The stability varies depending on the specimen.

#### 10.0 EXAMINATION PROCEDURE

##### 10.1 Preparation of specimens

- Sample feces using **Specimen Collection Container A**. See the instruction manual for the container.
- Shake the container sufficiently to dissolve the feces from the gutters of the stick.
- Leave the container at room temperature for 1 hour. When measuring on the next day, refrigerate at 2–8°C. Depending on the specimen, dissolution of transferrin from feces may be insufficient in 1 hour.

##### 10.2 Assay

- Prepared specimen solution

Prepared specimen solution	12 µL
Reagent 1	100 µL
Reagent 2	40 µL

Mix the reaction liquid and incubate at 37°C.

Formula to calculate the change of Absorbance (Abs) is (Am1–As1)–(Am2–As2).

Am1 : Abs of main wavelength at measurement point 1

As1 : Abs of sub wavelength at measurement point 1

Am2 : Abs of main wavelength at measurement point 2

As2 : Abs of sub wavelength at measurement point 2

main wavelength: 540 nm

sub wavelength: 660 nm

measurement point 1: 0 minutes

measurement point 2: 4.4 minutes

- Construct a calibration curve using **FIT Transferrin NS-Prime Calibrator** in place of the specimen solution. Read the human transferrin concentration in the specimen solution from the calibration curve.

All routine maintenance procedures defined in the instrument manual for the **Discrete Clinical Chemistry Analyzer NS-Prime** should be performed to obtain optimal performance.

#### 11.0 CONTROL PROCEDURE

It is recommended that each laboratory should use quality control materials routinely such as **FIT NS-Prime Control**, and should establish its own control ranges. Multilevel controls should be tested for each run. The human transferrin values obtained for the quality control materials should not fall repeatedly outside the control ranges established in each laboratory. If these control values fall repeatedly outside of the established control ranges, then proper instrument performance should be verified or perform recalibration.

#### 12.0 CALCULATION OF EXAMINATION RESULTS

Specimen values are calculated by the **Discrete Clinical Chemistry Analyzer NS-Prime**.

#### 13.0 INTERPRETATION OF RESULTS

The cut-off value widely used in Japan for mass-screening is less than 50 ng/mL (equivalent to 10 µg of human transferrin per 1 g of feces). Colorectal cancer is suspected if human transferrin is detected above that level.

<In-house data>

*Note : This value is indicative only and may differ from other published values because of differences in methods and in the population being studied. It is recommended that each laboratory establish its own cut-off value.*

#### 14.0 PERFORMANCE CHARACTERISTICS

The following performance data was obtained using the **Discrete Clinical Chemistry Analyzer NS-Prime**.

##### 14.1 Analytical performance characteristics

##### 14.1.1 Precision

Precision: CV ≤ 15%

##### 14.1.2 Interference

Some studies have been conducted to determine the levels of interference. Assay values of samples spiked with interference materials at the following concentrations ranged from 90% to 110% of the original value.

Conjugated and unconjugated bilirubin: 0–20,000 ng/mL

Ascorbic acid: 0–100,000 ng/mL

Glucose: 0–100,000 ng/mL

Bovine serum albumin: 0–100,000 ng/mL

Peroxidase: 0–100,000 ng/mL

Barium sulfate: 0–1,000,000 ng/mL

The cross-reactivities of the reagents with serum in animal species other than humans (bovine, swine, equine, ovine, leporine, and avian) were not been detected at a concentration of 0.4%.

##### 14.1.3 Correlation

**FIT Transferrin NS-Prime** assay was compared with the reference method on 100 specimens, whose transferrin concentrations were between 0 and 500 ng/mL. A linear regression analysis was performed and the following results were obtained:

y : **FIT Transferrin NS-Prime**

x : Reference method

r = 0.999

y = 1.022x – 0.479

#### 14.2 Diagnostic performance characteristics

##### 14.2.1 Analytical sensitivity

The difference in the amounts of change in absorbance between transferrin concentrations of 0 and 50 ng/mL is 0.05 or above.

##### 14.2.2 Analytical accuracy

Within 100±15% of known concentration in the measurement of a control specimen of known concentration.

#### 14.3 Measuring interval

##### 14.3.1 Assay range

- Upper limit : 500 ng/mL (equivalent to 100 µg human transferrin per 1 g of feces)
- Lower limit

limit of quantitation : 20 ng/mL (equivalent to 4 µg human transferrin per 1 g of feces)
limit of detection : 5 ng/mL (equivalent to 1 µg human transferrin per 1 g of feces)

#### 15.0 BIOLOGICAL REFERENCE INTERVALS

Less than 50 ng/mL (10 µg /g feces)

*Note : This value is indicative only and may differ from other published values because of differences in methods and in the population being studied. It is recommended that each laboratory establish its own local reference ranges.*

#### 16.0 LIMITATION OF THE EXAMINATION PROCEDURE












- Cuts, abrasions, and other skin lesions should be properly protected with an appropriate waterproof dressing, because blood contamination may influence measurements.
- Appearance changes, such as cloudiness and aggregation, in any of the reagents indicate the possibility of deterioration. Call your local dealer for advice.
- Mix the Reagent 1-Reagent 2 combined bottle thoroughly before each use, because colloidal gold particles may precipitate during storage. Mix it by slowly inverting to avoid creating bubbles. If bubbles occur, the instrument sensor will read an incorrect volume and the bottle will be deleted from the system. Pipette out any bubbles down to the level of the liquid.
- As with all assays, the results of this test can be influenced by factors present in some patients' specimens.
- For diagnostic purposes, the results obtained from this assay should always be used in combination with a clinical examination, patient medical history, and other findings.

6. Procedural directions must be followed exactly because any modification of the procedure may change the results.
7. Use of reagents, disposables, or spare parts other than those supplied by the authorized distributor may produce incorrect results.
8. Store the reagents according to the storage methods. Do not use them after the expiration date.
9. Use fresh feces.
10. This test should not be used to analyze specimens taken from a patient who is menstruating or who has hemorrhoids.
11. The test has not been validated for testing of patients with hemoglobinopathies.

#### 17.0 LITERATURE REFERENCES

1. Kazuyoshi Yamashita et al., The Usefulness of Fecal Transferring Test, Medical Journal of Sendai Red Cross Hospital, 19(1), p75–78, 2010.
2. Ichiro Hirata et al., Early Colorectal Cancer Screening-an Immunochemical Fecal Occult Blood Test Using Hb/Tf Simultaneous Assay, Stomach and Intestine, 45(5), p725–733, 2010.
3. Setsuko Kato et al., Basic Evaluation of Hemotect NS-Prime, an Automated Immunochemical Analyzer for Fecal Occult Blood Testing, J Clin Lab Inst Reag, 37(3), p371–377, 2014.
4. Wael L.L.Demian et al., Evaluation of the analytical performance of the novel NS-Prime system and examination of temperature stability of fecal transferrin compared with fecal hemoglobin as biomarkers in a colon cancer screening program, Practical Laboratory Medicine, 2, p29–36, 2015.
5. Yoshinori Takashima et al., Clinical benefit of measuring both haemoglobin and transferrin concentrations in faeces: demonstration during a large-scale colorectal cancer screening trial in Japan, Diagnosis, 2(1), p53–59, 2015.
6. Yoshinori Takashima et al., Corrigendum to: Clinical benefit of measuring both haemoglobin and transferrin concentrations in faeces: demonstration during a large-scale colorectal cancer screening trial in Japan, Diagnosis, 2(3), p199–200, 2015.
7. Taira Inagaki et al., Evaluation of the new automatic immunochemistry fecal occult blood analyzer "Hemo Tech NS-Prime", Japanese Journal of Medical Technology, 65(2), p222–228, 2016.

#### 18.0 SYMBOLS USED IN PRODUCT INSERTS AND ON LABELS

Symbols	Meanings of the symbols
	Expiry date (Used by...)
	Lot number
	Catalogue code
	Manufactured by
	Authorized EC representative
	Number of tests
	<i>In vitro</i> diagnostic medical device ( <i>In vitro</i> diagnostic use)
	Temperature limitation (store at...)
	See instruction for use
	Reagent 1
	Reagent 2

#### 19.0 DATE OF ISSUE OR REVISION

May 1, 2019